

## Influence of a novel organometallic Cu(II) complex on the photochemical activity of Photosystem II in spinach

Mehriban Shabanova<sup>1\*</sup>, Sergei Zharmukhamedov<sup>2</sup>, Suleyman Allakhverdiev<sup>1,2,3,4</sup>

<sup>1</sup>International Bionanotechnology Laboratory, Institute of Molecular Biology & Biotechnology, Ministry of Science and Education of the Republic of Azerbaijan, 11 Izzat Nabiyeu Str., AZ1143, Baku, Azerbaijan

<sup>2</sup>Institute of Basic Biological Problems, FRC PSCBR Russian Academy of Sciences, 142290, Pushchino, Moscow Region, Russia

<sup>3</sup>Controlled Photobiosynthesis Laboratory, K.A.Timiryazev Institute of Plant Physiology, Russian Academy of Sciences, 35 Botanicheskaya Str., 127276, Moscow, Russia

<sup>4</sup>Faculty of Engineering and Natural Sciences, Bahçeşehir University, 34349, Istanbul, Türkiye

\*For correspondence: mehriban\_shabanova@mail.ru

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**Photosynthesis is an attractive target for inhibitory compounds, both for the development of new herbicides and for advancing the understanding of photosynthetic processes. The synthetic [CuL<sub>2</sub>]Br<sub>2</sub> complex was studied for its inhibitory effect on the photosynthetic activity of photosystem II. It was demonstrated that a complex containing a benzothiazole group is an effective suppressor of photosynthetic activity.**

**Keywords:** Organometallic complexes, DCMU, OJIP, inhibitors

### INTRODUCTION

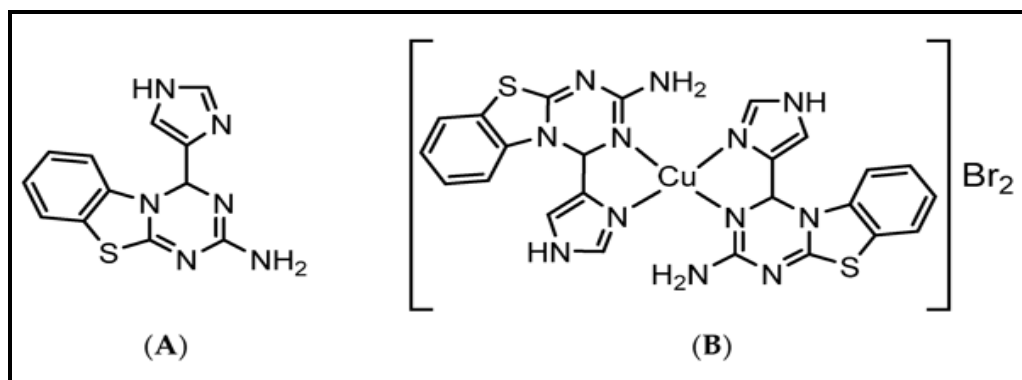
Herbicides remain the most effective method for controlling weeds. However, the repeated use of the same compounds leads to soil and water pollution, with chemical substances posing a risk of environmental damage (Vitek et al., 2017). Therefore, to mitigate environmental harm, special attention should be given to the development of new, effective, and selective compounds that act through different mechanisms. Photosynthesis is a complex process in which solar energy is converted into the energy of chemical bonds. As a vital process for all photosynthetic organisms, photosynthesis remains an attractive target for the application of inhibitory compounds (Zharmukhamedov et al., 2022). Currently, numerous chemical compounds are capable of inhibiting essential reactions in photosynthesis (Schütte et al., 2017).

However, compounds that affect only one of the metabolic pathways in plants are not very effective due to the evolving resistance of plants to their action. Therefore, the development of a universal inhibitor capable of suppressing a wide range of vital reactions represents a promising approach to addressing the resistance problem (Vass, 2012). In addition, chemical compounds can serve as tools for studying the mechanisms of photosynthetic reactions. Numerous exogenous

artificial electron donors and acceptors, as well as inhibitors, are widely used to separate the electron transport chain into distinct regions, allowing for their study without disrupting the thylakoid membrane with detergents.

Copper plays an important role in a variety of metabolic processes in plants, cyanobacteria, and algae (Yruea, 2005). Cu<sup>2+</sup> ions are essential for plant growth; however, high concentrations of Cu(II) exhibit the highest toxicity among heavy metal cations. It has been demonstrated that components of photosystem II (PSII) are more sensitive to the inhibitory effects of Cu than those of photosystem I (PSI) (Murakami et al., 2014). It is hypothesized that both the donor and acceptor sides of PSII are affected by Cu. Evidence suggests that Cu inhibits the activity of the PSII reaction center, enhances chlorophyll (Chl) degradation, and inhibits the activity of the water-oxidizing complex (WOC). Furthermore, Cu can disrupt PSII photochemical activity and alter the structure of thylakoid membranes, affecting the overall activity of the photosystems (Deng et al., 2014).

In this study, the [CuL<sub>2</sub>]Br<sub>2</sub> complex (where L = bis{4H-1,3,5-triazino[2,1-b]benzothiazole-2-amine, 4-(2-imidazole)}copper(II) bromide) was investigated for its inhibitory effect on the photochemical activity of spinach. Figure 1 shows the structure of the ligand (A) and the Cu(II)-complex (B).



**Fig. 1.** Ligand (L), 4H-1,3,5-triazino [2,1-b]benzothiazole-2-amine,4-(2-imidazole) (A), structure of [Cu(II)L<sub>2</sub>]Br<sub>2</sub> complex (B) (Zharmukhamedov et al., 2022).

## MATERIALS AND METHODS

**Isolation of PSII preparations.** PSII-enriched active thylakoid membrane fragments were extracted from leaves as described previously (Chiller & Dau, 2000). The PSII-containing membranes were suspended in medium (A) (50 mM MES-NaOH, pH 6.5, 300 mM sucrose, and 15 mM NaCl) and stored at  $-80^{\circ}\text{C}$ . The total chlorophyll concentration in the PSII-containing membranes was measured using 96% (v/v) ethanol (Arnon, 1949).

**Spectrophotometric measurements.** The absorption spectra of the [CuL<sub>2</sub>]Br<sub>2</sub> complex were measured using a standard quartz cell (Hellma, Müllheim, Germany) on a two-beam Shimadzu spectrophotometer (Shimadzu UV-1800, Shimadzu Europa GmbH, Duisburg, Germany) in the wavelength range of 200–700 nm at room temperature. The concentration of the [CuL<sub>2</sub>]Br<sub>2</sub> complex was 0.1 mM. Stock solutions of the [CuL<sub>2</sub>]Br<sub>2</sub> complex and DCMU (3-(3,4-dichlorophenyl)-1,1-dimethylurea) were prepared by dissolving in DMSO.

**Fast induction kinetics of chlorophyll fluorescence.** The fast induction kinetics of chlorophyll fluorescence was measured using a MULTI-COLOR-PAM fluorimeter (Heinz Walz GmbH, Pfullingen, Germany). All measurements were conducted at  $20^{\circ}\text{C}$  in a quartz cuvette (1 cm path length) at room temperature, following a dark adaptation period of at least 15 minutes. For the measurements, the chlorophyll concentration was  $4\ \mu\text{g mL}^{-1}$ .

## RESULTS AND DISCUSSION

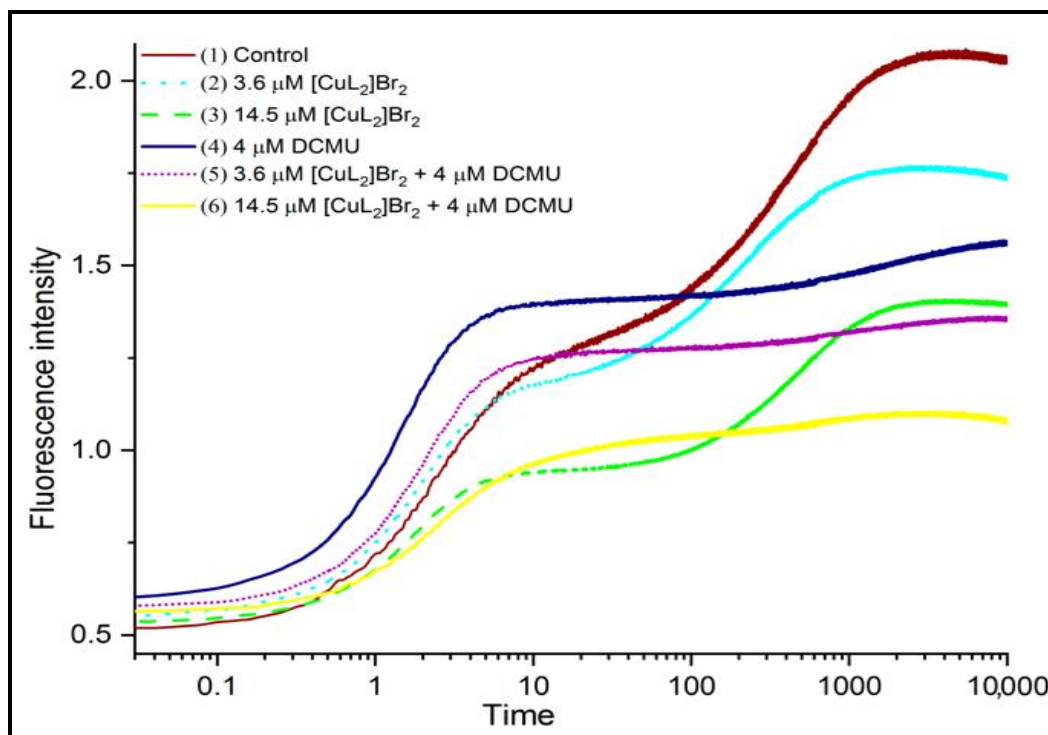
**Original OJIP kinetics.** The analysis of the OJIP test shown in Figure 2 reveals that all the investigated agents and their combinations lead to a

significant decrease in chlorophyll (Chl) fluorescence intensity, particularly at the  $F_M$  level. This results in a decrease in  $F_V$  (variable fluorescence). The  $F_V/F_M$  ratio is a widely used parameter that characterizes the quantum efficiency of the primary photochemical reaction in PSII (Kalaji et al., 2017, Pospíšil & Dau, 2002). Additionally, small increases in the  $F_0$  level were observed with  $3.6\ \mu\text{M}$  [CuL<sub>2</sub>]Br<sub>2</sub> (kinetics 2),  $14.5\ \mu\text{M}$  [CuL<sub>2</sub>]Br<sub>2</sub> (kinetics 3), and  $4\ \mu\text{M}$  DCMU (kinetics 4). An increase in the  $F_0$  level in the presence of DCMU has been previously observed in thylakoids and PSII-containing membranes (Pospíšil & Dau, 2000). In the presence of DCMU with both concentrations of [CuL<sub>2</sub>]Br<sub>2</sub> (kinetics 5 and 6), the increase in  $F_0$  is smaller than the increase observed with DCMU alone (kinetics 4).

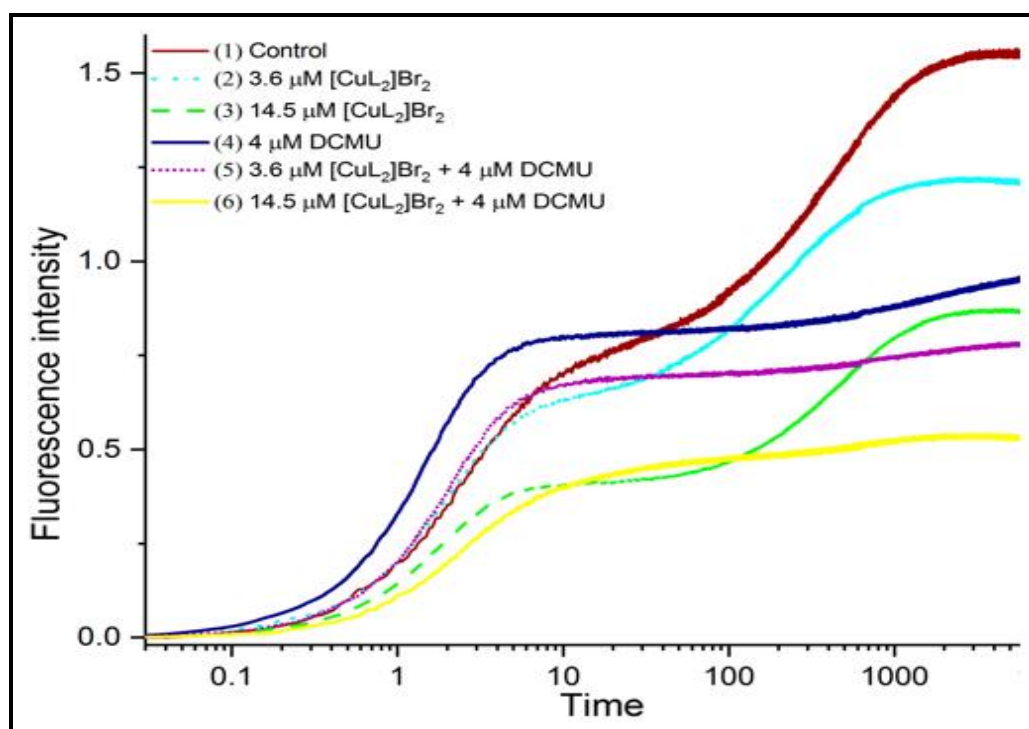
**Original OJIP Kinetics Normalized Relative to  $F_0$  ( $F_{0.02\text{ms}}$ ).** To simplify the analysis and more clearly represent the potential changes caused by the agents added to the control (without other additives), normalization is performed relative to the initial fluorescence level,  $F_0$ , using the value of  $F_0$  measured at  $20\ \mu\text{s}$  (Kalaji et al., 2017).

The original OJIP kinetics, normalized to  $F_0$ , are shown in Figure 3 as  $F_t - F_0$  versus time, where  $F_0$  represents the fluorescence at  $0.02\ \text{ms}$  and  $F_t$  represents the fluorescence at time  $t$ .

In the presence of both concentrations of [CuL<sub>2</sub>]Br<sub>2</sub>, a decrease in chlorophyll fluorescence intensity ( $F$ ) is observed throughout the entire OJIP kinetics. This decrease particularly affects the  $F_J$  level (2–3 ms), with a more noticeable effect observed at  $14.5\ \mu\text{M}$  [CuL<sub>2</sub>]Br<sub>2</sub>. The reduction in chlorophyll fluorescence is especially significant at the  $F_M$  level, where the kinetics for  $3.6\ \mu\text{M}$  and  $14.5\ \mu\text{M}$  [CuL<sub>2</sub>]Br<sub>2</sub> (kinetics 2 and 3) show a decrease compared with the control. The decrease in  $F_M$  is particularly pronounced at  $14.5\ \mu\text{M}$  [CuL<sub>2</sub>]Br<sub>2</sub> (kinetic 3). We will refer to these decreases in  $F$  (including  $F_J$  and  $F_M$ ) as the “effect of [CuL<sub>2</sub>]Br<sub>2</sub>.”



**Fig. 2.** Original OJIP kinetics, kinetic 1- control sample with PSII membranes in the absence any additions, kinetic 2 - 3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ ; kinetic 3 - 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ ; kinetic 4 - 4  $\mu\text{M}$  DCMU; kinetic 5 - 3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU; kinetic 6 - 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU.



**Fig. 3.** OJIP kinetics normalized relative to  $F_{0.02\text{ms}}$ , kinetic 1- control sample with PSII membranes in the absence any additions, kinetic 2 - 3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ ; kinetic 3 - 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ ; kinetic 4 - 4  $\mu\text{M}$  DCMU ; kinetic 5 - 3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU; kinetic 6 - 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU.

We have determined the percentage change in  $F_M$  from the control based on the results obtained in Figure 2. The decrease in  $F_M$  for each condition,

compared to the control, is as follows:

- Kinetic 2 (3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ : 22%
- Kinetic 3 (14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ : 45%

- Kinetic 4 (4  $\mu\text{M}$  DCMU): 38%
- Kinetic 5 (3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU): 50%
- Kinetic 6 (14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU): 66%

Thus, these experimental data indicate that 22% and 45% of the total PSII-containing membranes (kinetic 1 and kinetic 2) are no longer able to carry out the photochemical reduction of the corresponding components on the acceptor side of PSII. This is due to the specific suppressive effect of  $[\text{CuL}_2]\text{Br}_2$  on the components responsible for either charge separation or electron transfer from the donor side to the components of PSII.

In addition,  $F_M$  is suppressed in the presence of 4  $\mu\text{M}$  DCMU and, particularly, in the presence of its combinations with both concentrations of  $[\text{CuL}_2]\text{Br}_2$ . Moreover, when 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  is combined with 4  $\mu\text{M}$  DCMU, an almost synchronous decrease in chlorophyll fluorescence intensity ( $F$ ) is observed throughout the entire OJIP kinetics. In the presence of DCMU (without  $[\text{CuL}_2]\text{Br}_2$ ), we observed an increase in the  $F_J$  peak to the so-called  $F_M$  peak (DCMU effect). In the presence of DCMU, the entire amount of  $Q_A$  in the sample is reduced, resulting in an increase in the  $J$  peak to its maximum possible level. At the same time, the  $F_M$  value decreases to 62% of the control  $F_M$  value. The changes in OJIP kinetics observed in the presence of both DCMU and  $[\text{CuL}_2]\text{Br}_2$  are similar to those recorded in the presence of only DCMU (the “DCMU effect”), but the percentage decrease is even more significant over the entire OJIP kinetics (the “[ $\text{CuL}_2]\text{Br}_2$  effect”). This decrease is especially pronounced in kinetic 6 (14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$  + 4  $\mu\text{M}$  DCMU).

## DISCUSSION

**Main inhibitory effect of  $[\text{CuL}_2]\text{Br}_2$  on OJIP transient.** The most pronounced and, therefore, undoubtedly the primary effect of the  $[\text{CuL}_2]\text{Br}_2$  complex on the photochemical activity of PSII-containing membranes is the synchronous decrease in fluorescence intensity along the entire JIP kinetics (Figures 2 and 3, kinetics 2, 3). At a concentration of 3.6  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ , 22% of PSII-containing membranes are completely excluded from the kinetics, but at 14.5  $\mu\text{M}$   $[\text{CuL}_2]\text{Br}_2$ , 45% of PSII-containing membranes are already fully inhibited (in the absence of DCMU). Importantly,  $[\text{CuL}_2]\text{Br}_2$  demonstrates this effect on PSII even in the presence of DCMU, with an efficiency comparable to that observed in its absence. Based on these data, we can conclude that the primary effect of  $[\text{CuL}_2]\text{Br}_2$  on PSII is independent of

DCMU. The reduction of  $F_M$  observed at both concentrations of  $[\text{CuL}_2]\text{Br}_2$ , in the presence and absence of DCMU, suggests that PSII inhibition by  $[\text{CuL}_2]\text{Br}_2$  may occur via the same mechanism of action in both cases. The fact that  $[\text{CuL}_2]\text{Br}_2$  inhibits  $F_M$  regardless of the presence of DCMU can be explained by the hypothesis that the site of action of  $[\text{CuL}_2]\text{Br}_2$  on PSII precedes the site of action of DCMU. The simultaneous decrease in the  $F_M$  intensity of chlorophyll in the OJIP kinetics, which increases with the concentration of  $[\text{CuL}_2]\text{Br}_2$ , may be the result of the destruction of the donor side or the reaction center (RC) of PSII. Similar results in the OJIP test were observed when the donor side of PSII became inactive (Zharmukhamedov et al., 2022).

## CONCLUSIONS

According to the results of this study,  $[\text{CuL}_2]\text{Br}_2$  exerts various interesting effects on different regions of PSII. The results demonstrate that the primary effect of  $[\text{CuL}_2]\text{Br}_2$  on PSII is likely associated with the inhibition of the activity of the PSII reaction center.  $[\text{CuL}_2]\text{Br}_2$  effectively reduces the  $F_M$  value both in the absence and presence of DCMU. The obtained data may be useful for developing effective herbicides for agricultural applications.

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## REFERENCES

- Arnon D.I.** (1949) Copper enzymes in isolated chloroplasts. Polyphenol oxidase in *Beta vulgaris*. *Plant. Physiol.*, **24**: 1-17.
- Chiller H., Dau H.** (2000) Preparation protocols for high-activity photosystem II membrane particles of green algae and higher plants, pH dependence of oxygen evolution and comparison of the  $S_2$ -state multiline signal by X-band EPR spectroscopy. *J. Photochem. Photobiol. B*, **55**:

- 138-144
- Deng C., Pan X., Wang S., Zhang D.** (2014) Cu<sup>2+</sup> inhibits photosystem II activities but enhances photosystem I quantum yield of *Microcystis aeruginosa*. *Biol. Trace Elem. Res.*, **160**(2): 268-275.
- Kalaji M.H., Goltsev V.N., Zuk-Golaszewska K., Zivcak M., Brestic M.** (2017) Chlorophyll fluorescence, understanding crop performance. *CRC Press: Boca Raton, FL, USA; ISBN 978-1-4987-6449-0*.
- Murakami K., Tsubouchi R., Fukayama M., Yoshino M.** (2014) Copper dependent inhibition and oxidative inactivation with affinity cleavage of yeast glutathione reductase. *Biometals*, **27**: 551-558.
- Pospíšil P., Dau H.** (2000) Chlorophyll fluorescence transients of photosystem ii membrane particles as a tool for studying photosynthetic oxygen evolution. *Photosynth. Res.*, **65**: 41-52.
- Pospíšil P., Dau H.** (2002) Valinomycin sensitivity proves that light-induced thylakoid voltages result in millisecond phase of chlorophyll fluorescence transients. *Biochim. Biophys. Acta Bioenerg.*, **1554**: 94-100.
- Schütte G., Eckerstorfer M., Rastelli V., Reichenbecher W., Restrepo-Vassalli S., Ruohonen-Lehto M., Saucy A.G.W., Mertens M.** (2017) Herbicide resistance and biodiversity: agronomic and environmental aspects of genetically modified herbicide-resistant plants. *Environ. Sci. Eur.*, **29**: 5.
- Vass I.** (2012) Molecular mechanisms of photodamage in the photosystem ii complex. *Biochim. Biophys. Acta Bioenerg.*, **1817**: 209-217.
- Vítek P., Novotná K., Hodaňová P., Rapantová B., Klem K.** (2017) Detection of herbicide effects on pigment composition and PSII photochemistry in *Helianthus annuus* by Raman spectroscopy and chlorophyll a fluorescence. *Spectrochim Acta A Mol Biomol. Spectrosc.*, **170**: 234-241.
- Yruea I.** (2005) Copper in plants. *Braz. J. Plant Physiol.*, **17**(1): 145-156.
- Zharmukhamedov S.K., Shabanova M.S., Rodionova M.V., Huseynova I.M., Karacan M.S., Karacan N., Asik K.B., Kreslavski V.D., Alwasel S., Allakhverdiev S.I.** (2022) Effects of novel photosynthetic inhibitor [CuL<sub>2</sub>]Br<sub>2</sub> complex on photosystem ii activity in spinach. *Cells*, **11** (7): 2680.

## İspanaqda fotosistem II-nin fotokimyəvi aktivliyinə yeni Cu(II) orqanometalik kompleksinin təsiri

Mehriban Şabanova<sup>1</sup>, Sergey Jarmuxamedov<sup>2</sup>, Süleyman Allahverdiyev<sup>1,2,3,4</sup>

<sup>1</sup>Azərbaycan Respublikası Elm və Təhsil Nazirliyi Molekulyar Biologiya və Biotexnologiyalar İnstitutunun Bionanotexnologiya beynəlxalq laboratoriyası, Bakı, Azərbaycan

<sup>2</sup>Rusiya Elmlər Akademiyası Biologiyanın Fundamental Problemləri İnstitutu, Puşino, Rusiya

<sup>3</sup>Rusiya Elmlər Akademiyası K.A. Timiryazev adına Bitki Fiziologiyası İnstitutu İdarəolunan Fotobiosintez laboratoriyası, Moskva, Rusiya

<sup>4</sup>Bahçeşehir Universitetinin Mühəndislik və təbiət elmləri fakültəsi, İstanbul, Türkiyə

Fotosintez günəşdən gələn enerji hesabına bitkilərin böyüməsini və inkişafını təmin edir və bitki hüceyrələrinin malik olduğu unikal xüsusiyyətidir. Fotosintetik aparatın (FA) əsas komplekslərindən biri olan fotosistem II (FSII) suyu elektronlara, protonlara və oksigenə parçalayır. Cu(II)<sup>+</sup> aqua ionlarının kompleksləri və liqandları FSII-nin fotosintetik fəaliyyətinə tormozlayıcı təsir göstərmək üçün tədqiq edilmişdir. Müxtəlif FSII komponentlərində Cu<sup>2+</sup> aqua ionlarının müxtəlif təsir saytları və effektləri təyin edilmişdir.

**Açar sözlər:** Üzvü-metal komplekslər, DCMU, OJIP, inhibitorlar

### ORCIDS:

Mehriban Shabanova: <https://orcid.org/0009-0000-0502-4085>

Sergey Zharmukhamedov: <https://orcid.org/0000-0002-8742-1072>

Suleyman Allakhverdiev: <https://orcid.org/0000-0002-0452-232X>